

## Evaluation and quantification of digestive amylase enzyme and its relation with productive and economic traits of multivoltine silkworm genetic resources of India

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### Abstract

Amylase is one of the key enzymes involved in the digestion and carbohydrate metabolism in insects. There is wide genetic variability in the activity of digestive amylase among the silkworm genetic resources of varying geographical origin and yield potential. The digestive amylase is generally very high in the non diapausing multivoltine silkworm races than that of the diapausing bivoltine stocks. Therefore in the present study an attempt has been made to evaluate and quantify the digestive amylase enzyme activity in the 73 multivoltine silkworm genetic resources conserved at Central Sericultural Germplasm Resources Centre, Hosur which is the only centre where all the multivoltine silkworm genetic resources were conserved following standard protocols. The study has been also carried out with the objective of revealing the quantum of variability of the amylase enzyme activity in the multivoltine silkworm genetic resources of the varying geographical origin and their association with that of the productive and economic traits. The study revealed that significant variability in the quantity of amylase 1600 µg to 2250 µg among the 73 MV silkworm genetic resources evaluated. Positive correlations in some of the traits like weight of 10 larvae, effective rate of rearing by number and single cocoon weight and cocoon yield per 100 dfls were observed. Some of the traits like total larval duration, effective rate of rearing by number, pupation and single shell weight showed negatively correlation. Significant negative correlations were noticed in fecundity, hatching, fifth age larval duration and shell ratio.

**Keywords:** amylase, silkworm genetic resources, correlation, economic traits

### 1. Introduction

Mulberry silkworm *Bombyx mori* L. feeds on the leaves of *Morus spp.* for its food and to grow and develop and completes its lifecycle. During the only active feeding stage it consumes large quantity of mulberry leaves for completion of its biological activities and processes. As mulberry a perennial tree plant grown although the year it is being influenced by the by the seasonal changes and also the inputs given to the plant. Probably most of the study of characteristics of an insect digestive enzyme has been made on amylase of silkworm (Mori, 1930; Ito *et al.*, 1962) [19, 16]. Kanekatsu (1972) [17] purified amylase of the digestive fluid and studied properties of amylase in *Bombyx mori*. Banano *et al.* (1984) [4] reported the sex dimorphic and development changes in amylase of haemolymph of *Bombyx mori*. Amylases obtained from different origins have been characterized by Baker (1989) [3] and Bhawane and Bhanot (1989) [5]. In silkworm studies on the different amylase activity among silkworm strains and relationship between amylase activity of the larval digestive juice and economic characters has been done (Hirata, 1971; 1974) [13-14]. Amylase one of the key enzymes involved in the digestion and carbohydrate metabolism in insects (Horie and Watanabe, 1983) [15] and particularly carbohydrates available in the form of starch in mulberry leaves (Chatterjee *et al.*, 1989) [7]. Abraham *et al.* (1992) [1] identified different series of amylases in both diapausing and non diapausing strains of *Bombyx mori*. The wide genetic variability in the activity of digestive amylase has been indicated when large number of silkworm stocks of wide geographical origin and yield status at different research centres and most of the high yielding,

diapausing stocks and low amylase activity while most of the low yielding non diapausing stocks showed high amylase activity. In general the activities of amylase in multivoltine races were almost two fold higher than the bivoltine races of silkworms (Chatterjee *et al.*, 1992) [9].

The information on the importance of digestive amylase activity in yield and biochemical parameters, of late molecular markers facilitated selection using isozyme / DNA markers is being widely explored and successfully used in the breeding of crop plants and livestock for the management of target traits (Young and Tanksley, 1989; Tankslet and Nelson, 1996; Visscher *et al.*, 1996; Fulton *et al.*, 1997) [27, 23, 26, 12]. The development profiles of amylase enzyme, its genetic variability and activity in the mulberry silkworm its significance and the correlation between yield and biochemical parameters were investigated by (Chatterjee *et al.*, 1989; 1990; 1992; 1993) [7-10]. Patnaik and Datta (1995) [22] studied the type of amylase and their activity in the bivoltines and multivoltines strains. In view of the wide divergence in terms of activity and isozyme polymorphism and its role in better digestibility and close association with survival, digestive amylase isozyme was chosen as a marker in silkworm breeding for improving the potential of productive breeds (Ashwath *et al.*, 2001) [2]. The function of the digestive fluid amylase is related to the digestion of starch contained in the food plant - mulberry (Kanekatsu, 1972) [17]. Muniv *et al.* (2011) [21] studied the characteristics of midgut amylase and made a comparative analysis in multivoltine races Nistari and Kolar Gold.

Therefore in the present study an attempt has been made to screen the amylase activity in the different multivoltine

silkworm germplasm resources conserved at CSGRC, Hosur which will indicate the extent of variability in the quantity of amylase among the 73 germplasm stocks of different geographical origins, productivity and economic important traits which can be used in future studies. Also the correlation between quantum of amylase and productivity, survival and also the deviations if any among the multivoltine genetic stocks of the contry.

## 2. Materials and Methods

The studies were undertaken at the Central Sericultural Germplasm Resources Centre, Hosur. Seventy three multivoltine silkworm accessions conserved at the centre were utilized for the investigations. Sixty three of these

accessions are from different geographical regions of the country and 10 accessions were of exotic origin (Table-1). The accessions evaluated possess greater variability in terms of morphological and economic traits and also comprising many native breeds, land races, breeds under current use, breeding resources materials and also few obsolete breeds. The dfls of these accessions are conserved by cold preserving the eggs for 35 days under 5°C and thereafter releasing them and incubating at 25°C and 80 % RH. On attaining pin head stage they will be composited by taking 50 eggs from each of the 20 dfls preserved at two different cold storages and composite populations will be brushed after black boxing the composited eggs. (Thangavelu *et al.*, 1997; 2000)<sup>[24, 25]</sup>.

**Table 1:** Details of multivoltine accessions vis a vis their morphological traits.

Accession No.	Race Name	Origin	Class	Egg colour	Yolk Colour	Larval pattern	Cocoon colour	Cocoon shape
BMI-0001	PURE MYSORE	India/Karnataka	O(BCU)	Dull white	Light yellow	Plain	LGY	Spindle
BMI-0002	SARUPAT	India/Assam	O(OB)	Light yellow	Creamish white	Plain	Creamy white	SA
BMI-0003	MORIA	India/Assam	O(OB)	FYGT	Light yellow	Plain	Creamy white	SA
BMI-0004	TAMILNADU WHITE	India/Tamil Nadu	E(OB)	White	Creamish white	Marked	White	SA
BME-0005	C.NICHI	Japan	O(OB)	White	Creamish white	Marked	White	Constricted
BMI-0006	HOSA MYSORE	India/Karnataka	E(BCU)	FYGT	Yellow	Plain	Greenish yellow	Elongated oval
BMI-0007	MYSORE PRINCESS	India/Karnataka	E(OB)	White	Creamish white	Plain	White	Oval
BMI-0008	KOLAR GOLD	India/Karnataka	E(OB)	White	Creamish white	Plain	White	Oval
BMI-0009	KOLLEGAL JAWAN	India/Karnataka	E(OB)	White	Creamish white	Plain	White	Oval
BMI-0010	MY-1	India/Karnataka	E(BCU)	FYGT	Light yellow	Plain	LGY	Elongated
BMI-0011	P2D1	India/Karnataka	E(BCU)	FYGT	Yellow	Marked	Greenish yellow	Elongated oval
BME-0012	RONG DAIZO	China	E(OB)	FYGT	Yellow	Marked	Greenish yellow	SA
BME-0013	GUANGNONG PLAIN	China	E(OB)	White	Creamish white	Plain	Creamy white	Oval
BMI-0014	OS-616	India/West Bengal	E(OB)	FYGT	Yellow	Marked	Yellow	Oval
BME-0015	RAJ	Bangladesh	O(OB)	FYGT	Light yellow	Marked	Creamy white	Spindle
BMI-0016	G	India/West Bengal	E(BCU)	FYGT	Yellow	Marked	Golden yellow	Oval
BMI-0017	NISTARI	India/West Bengal	O(BCU)	FYGT	Yellow	Marked	Golden yellow	Spindle
BMI-0018	NISTARI(M)	India/West Bengal	E(BM)	FYGT	Yellow	Marked	Golden yellow	SB
BMI-0019	NISTARI(P)	India/West Bengal	E(BM)	FYGT	Yellow	Plain	Golden yellow	SB
BMI-0020	ZPN(SL)	India/West Bengal	E(BM)	Faint yellow	Creamish white	Mixed	Creamy white	Spindle
BMI-0021	CB5	India/West Bengal	E(BCU)	FYGT	Yellow	Marked	Golden yellow	Oval
BMI-0022	KW2	India/West Bengal	E(BM)	Creamish white	Creamish white	Plain	Creamy white	Elongated oval
BMI-0023	M2	India/West Bengal	E(BM)	FYGT	Yellow	Marked	Golden yellow	Oval
BMI-0024	A23	India/West Bengal	E(BM)	FYGT	Yellow	Marked	Golden yellow	Oval
BMI-0025	A25	India/West Bengal	E(BM)	FYGT	Yellow	Marked	Golden yellow	Elongated oval
BMI-0026	OVAL	India/West Bengal	E(BM)	FYGT	Yellow	Marked	Golden yellow	Oval
BMI-0027	O	India/West Bengal	E(OB)	FYGT	Yellow	Marked	Golden yellow	Oval
BMI-0028	M83(C)	India/West Bengal	E(BM)	FYGT	Yellow	Marked	Golden yellow	Oval
BMI-0029	B	India/West Bengal	E(BM)	FYGT	Yellow	Marked	Golden yellow	Elongated
BME-0030	GNM	China	E(OB)	White	Yellow	Marked	White	Oval
BMI-0031	A14DY	India/West Bengal	E(BM)	FYGT	Yellow	Marked	Golden yellow	Oval
BMI-0032	A4e	India/West Bengal	E(OB)	FYGT	Yellow	Marked	Greenish yellow	Elongated oval
BMI-0033	PA12	India/Karnataka	E(BM)	FYGT	Yellow	Plain	Greenish yellow	Elongated oval
BMI-0034	AP12	India/Karnataka	E(BM)	FYGT	Yellow	Marked	Greenish yellow	Elongated oval
BMI-0035	A13	India/Karnataka	E(BM)	FYGT	Yellow	Plain	Greenish yellow	Elongated oval
BMI-0036	PMX	India/Karnataka	E(BM)	FYGT	Yellow	Plain	LGY	Elongated oval
BMI-0037	PMS2	India/Karnataka	E(BM)	FYGT	Yellow	Plain	LGY	Elongated oval
BMI-0038	MU-1	India/Karnataka	E(BM)	FYGT	Yellow	Plain	LGY	Elongated oval
BMI-0039	MU-11	India/Karnataka	E(BM)	FYGT	Yellow	Plain	LGY	Elongated oval
BMI-0040	WAI-1	India/Maharashtra	E(BM)	FYGT	Yellow	Marked	LGY	Elongated oval
BMI-0041	WAI-4	India/Maharashtra	E(OB)	FYGT	Bright yellow	Marked	Yellow	Oval
BMI-0042	MY23	India/Karnataka	E(BM)	FYGT	Yellow	Plain	Greenish yellow	Elongated oval
BMI-0043	MW13	India/Karnataka	E(BM)	White	Creamish white	Plain	White	Oval
BMI-0044	MHMP(W)	India/Karnataka	E(BM)	White	Creamish white	Plain	White	Elongated oval
BMI-0045	MHMP(Y)	India/Karnataka	E(BM)	FYGT	Yellow	Plain	Greenish yellow	Elongated oval
BMI-0046	P4D3	India/Karnataka	E(OB)	FYGT	Yellow	Marked	Greenish yellow	Elongated oval

BME-0047	NISTID(Y)	Bangladesh	E(OB)	FYGT	Yellow	Plain	Golden yellow	SA
BME-0048	NISTID(W)	Bangladesh	E(OB)	White	Creamish white	Plain	White	SA
BME-0049	NK4	Japan	E(OB)	Light yellow	Light yellow	Marked	Yellow	SA
BME-0050	CAMBODG	Japan	O(OB)	Creamish white	Light yellow	Marked	Yellow	SA
BME-0052	DAIZO	China	O(OB)	FYGT	Yellow	Marked	Greenish yellow	Spindle
BMI-0053	LMP	India/West Bengal	E(BM)	FYGT	Yellow	Marked	Golden yellow	SA
BMI-0054	DMR	India/West Bengal	E(BM)	FYGT	Yellow	Marked	Golden yellow	Oval
BMI-0055	LMO	India/West Bengal	E(BM)	FYGT	Yellow	Marked	Golden yellow	Oval
BMI-0056	MY1(SL)	India/Karnataka	E(BM)	FYGT	Yellow	Mixed	LGY	Elongated
BMI-0057	PM(SL)	India/Karnataka	E(BM)	FYGT	Yellow	Mixed	LGY	Elongated oval
BMI-0058	BL23	India/Karnataka	E(BCU)	Light yellow	FYGT	Plain	Greenish yellow	ENC
BMI-0059	BL24	India/Karnataka	E(BCU)	Light yellow	FYGT	Plain	Greenish yellow	ENC
BMI-0060	MU303	India/Karnataka	E(BM)	Light yellow	FYGT	Plain	Greenish yellow	ENC
BMI-0061	MU520	India/Karnataka	E(BM)	Light yellow	Colourless	Plain	White	Oval
BMI-0062	MU10	India/Karnataka	E(BM)	Light yellow	Colourless	Plain	White	Oval
BMI-0063	TW x SK6 x SK1	India/Tamil Nadu	E(BM)	Light yellow	Colourless	Plain	White	DB
BMI-0064	SK6 x SK1 x TW	India/Tamil Nadu	E(BM)	Light yellow	Colourless	Plain	White	DB
BMI-0065	BL43	India/Karnataka	Evolved	Light yellow	Colourless	Plain	Greenish yellow	Elongated
BMI-0066	APM-1	India/Andhra Pradesh	Evolved	Yellow	Colourless	Plain	Greenish yellow	Oval
BMI-0067	SLKSPM	India/Karnataka	Evolved	Light yellow	Colourless	Mixed	Greenish yellow	Elongated
BMI-0068	M12(W)	India/West Bengal	Evolved	Light yellow	Colourless	Plain	White	Spatulate
BMI-0069	M15	India/West Bengal	Evolved	Light yellow	Colourless	Plain	White	Spatulate
BMI-0070	M6DP(C)	India/West Bengal	Evolved	Light yellow	Colourless	Plain	Chrome yellow	Spatulate
BMI-0071	M6DP(C)Green	India/West Bengal	Evolved	Light yellow	Light yellow	Plain	White	Spatulate
BMI-0072	M6M81	India/West Bengal	Evolved	Light yellow	Light yellow	Plain	Chrome yellow	Oval
BMI-0073	BL-67	India/Karnataka	Evolved	Light yellow	Colourless	Plain	Greenish yellow	Oval
BMI-0074	MH-1	India/Karnataka	Evolved	Light Yellow	Light yellow	Marked	Creamy White	Spindle

The neonates were brushed upon hatching and were reared at standard rearing conditions (Krishnaswami, 1978) [18]. After third moult 250 larvae were counted and kept in 3 replications. The economic parameters like fecundity, hatching percentage, 10 larval weight, total larval duration,

fifth age larval duration, effective rate of rearing by number, effective rate of rearing by weight, pupation rate, single cocoon weight, single shell weight, shell ratio and cocoon yield/ 100 dfls were recorded (Table-2) for analysis.

**Table 2:** The growth and economic traits of 74 multivoltine genetic resources alongwith Amylase quantity.

Accession No.	Fecundity (No.)	Hatching (%)	Weight of 10 larvae g.	Total larval duration h.	Fifth age larval duration h.	ERR by No.	ERR By Wt. Kg.	Pupation (%)	Single Cocoon weight g.	Single Shell Weight g.	Shell Ratio (%)	Yield / 100 dfls kg.	Amylase quantity in µg /10µl /hour
BMI-0001	432	94.0	29.89	576	144	9612	10.65	91.29	1.01	0.14	13.54	42.59	2050
BMI-0002	376	90.8	31.50	504	120	8679	10.05	85.12	0.93	0.12	13.56	40.10	2103
BMI-0003	428	93.2	29.00	500	116	9097	11.10	88.28	0.93	0.14	16.10	44.37	2000
BMI-0004	342	93.6	27.00	492	108	9254	10.25	87.67	0.94	0.13	13.83	40.88	2100
BME-0005	438	93.6	25.50	492	108	9379	10.10	92.37	0.86	0.12	14.14	40.32	1900
BMI-0006	421	89.0	26.10	492	108	9780	10.25	96.33	0.90	0.13	14.38	40.88	1875
BMI-0007	436	92.6	31.35	504	120	9473	12.45	92.66	1.06	0.18	16.56	49.80	1950
BMI-0008	387	91.6	27.44	504	120	9791	10.00	94.94	0.90	0.13	14.35	40.04	1975
BMI-0009	381	92.8	28.98	504	120	9547	11.10	93.38	0.92	0.13	13.79	22.32	2050
BMI-0010	402	91.3	24.02	504	120	9733	10.15	96.39	0.91	0.11	12.22	40.66	2000
BMI-0011	335	91.0	26.60	504	120	9720	10.55	94.34	0.92	0.13	14.75	42.25	2250
BME-0012	436	94.5	26.56	576	144	9707	10.95	96.27	0.95	0.14	15.19	43.78	2150
BME-0013	390	87.6	24.74	536	116	9712	10.45	94.42	0.76	0.10	12.89	41.93	1900
BMI-0014	345	90.4	22.71	516	132	9804	11.30	97.13	0.92	0.14	15.29	45.04	1975
BME-0015	441	93.2	26.41	516	132	9567	10.15	93.32	0.80	0.11	13.99	40.60	1650
BMI-0016	295	92.2	26.73	500	116	9683	10.60	93.50	0.93	0.13	14.66	42.46	2053
BMI-0017	342	95.3	25.86	500	116	9747	9.80	95.44	0.86	0.11	13.48	39.19	1950
BMI-0018	409	95.0	24.53	500	116	9689	10.00	95.76	0.82	0.11	14.02	40.07	2000
BMI-0019	449	93.5	25.69	500	116	9784	10.25	95.48	0.84	0.11	13.78	41.07	1950
BMI-0020	353	90.0	23.93	528	124	9686	10.05	95.61	0.80	0.12	15.10	40.20	2000
BMI-0021	397	91.6	24.31	500	96	9667	9.80	95.33	0.83	0.12	14.62	39.20	1953
BMI-0022	306	89.9	24.83	504	100	9705	11.75	95.79	0.92	0.14	15.28	46.99	1975
BMI-0023	469	87.0	25.36	528	124	9640	10.55	94.73	0.95	0.16	16.73	42.16	2050
BMI-0024	520	95.9	27.93	528	124	9563	10.75	89.40	0.93	0.15	16.44	42.95	1900
BMI-0025	424	92.9	29.57	516	112	9778	10.85	94.69	0.91	0.14	15.16	43.36	2103
BMI-0026	345	92.5	29.85	516	112	9484	10.05	91.52	0.98	0.15	15.83	40.30	2050
BMI-0027	365	92.3	25.64	528	144	9876	10.95	97.27	1.05	0.15	14.52	43.72	2250

BMI-0028	306	89.8	26.46	500	116	9734	9.90	94.99	0.80	0.12	15.36	39.67	2050
BMI-0029	361	92.2	25.11	500	116	9700	10.10	95.33	0.77	0.11	14.48	40.33	2050
BME-0030	328	90.0	29.73	516	132	9390	11.80	92.96	0.89	0.13	14.93	47.13	2103
BMI-0031	369	91.9	25.36	492	108	9752	10.05	96.19	0.76	0.11	15.05	40.19	1950
BMI-0032	305	90.1	24.87	492	108	9650	9.75	94.17	0.98	0.15	15.25	39.13	1875
BMI-0033	376	88.9	28.16	486	102	9800	10.00	96.87	0.97	0.13	14.07	39.92	2150
BMI-0034	424	94.9	28.20	486	102	9718	10.05	93.89	0.99	0.16	15.99	40.17	2103
BMI-0035	367	92.9	27.09	486	102	9768	9.95	94.71	0.91	0.12	13.26	39.80	2050
BMI-0036	411	88.6	26.16	486	102	9669	10.25	94.97	0.94	0.13	13.86	41.14	2103
BMI-0037	468	93.3	28.42	486	102	9746	10.05	96.06	1.01	0.10	11.04	40.18	1953
BMI-0038	396	87.2	30.80	486	102	9666	9.95	94.87	0.92	0.13	14.52	39.85	2150
BMI-0039	482	91.3	28.31	486	102	9677	10.15	95.20	0.95	0.14	15.02	40.62	2050
BMI-0040	450	90.4	28.38	500	116	9699	10.20	94.99	0.98	0.13	13.83	40.84	2050
BMI-0041	454	93.4	28.93	500	116	9679	10.65	96.25	1.02	0.15	14.64	42.64	2100
BMI-0042	357	93.0	20.48	500	116	9619	10.10	93.00	0.89	0.13	14.94	40.29	2150
BMI-0043	445	93.2	28.65	500	116	9544	11.10	94.14	1.07	0.17	15.72	44.49	2103
BMI-0044	421	95.7	28.99	492	108	9747	10.50	95.70	1.03	0.15	14.57	42.07	1953
BMI-0045	358	89.1	28.71	492	108	9776	10.35	95.66	0.99	0.14	14.38	41.31	2100
BMI-0046	441	91.0	29.20	492	108	9691	10.55	95.07	1.09	0.15	13.32	42.19	2050
BME-0047	315	88.7	23.81	492	108	9583	10.45	94.29	0.81	0.12	14.59	41.88	2100
BME-0048	346	91.3	30.91	492	108	9721	10.35	93.31	0.89	0.14	16.13	41.45	1980
BME-0049	393	90.3	29.02	492	108	9830	10.30	97.10	0.83	0.12	14.49	41.27	2100
BME-0050	433	91.9	27.48	492	108	9657	10.80	95.12	0.92	0.13	13.78	43.21	2050
BME-0052	460	93.3	27.41	528	144	9673	10.40	95.70	0.82	0.13	16.27	41.51	1650
BMI-0053	322	86.9	27.78	500	116	9753	10.10	95.89	0.85	0.12	14.41	40.34	2050
BMI-0054	366	90.3	25.54	500	116	9471	10.25	88.82	0.91	0.14	16.04	40.97	2100
BMI-0055	474	92.2	29.20	500	116	9648	10.45	95.37	1.00	0.15	15.02	41.82	2100
BMI-0056	400	93.2	25.55	500	116	9696	10.20	94.43	0.96	0.14	14.54	40.83	2000
BMI-0057	359	87.5	25.65	500	116	9707	10.55	96.00	0.89	0.13	14.91	42.17	2100
BMI-0058	460	90.7	27.39	486	102	9817	10.90	96.71	0.91	0.13	14.05	43.67	1950
BMI-0059	471	90.8	28.57	486	102	9635	11.10	95.00	0.90	0.13	14.97	44.31	1950
BMI-0060	360	90.6	27.33	486	102	9792	10.60	95.00	0.94	0.14	15.24	42.39	1950
BMI-0061	384	91.9	29.88	516	132	9699	10.45	94.65	1.07	0.17	16.15	41.65	1950
BMI-0062	381	89.0	28.84	500	116	9763	10.25	94.84	0.96	0.15	16.22	41.10	1953
BMI-0063	363	92.2	30.18	486	102	9766	10.60	94.99	0.81	0.10	12.88	42.39	1950
BMI-0064	391	93.3	25.98	486	102	9833	11.05	96.63	0.79	0.11	13.89	44.26	1875
BMI-0065	450	94.5	31.23	516	132	9767	8.45	96.14	0.98	0.17	17.51	33.91	1900
BMI-0066	378	92.5	31.77	516	132	9712	10.95	95.88	1.07	0.17	16.43	43.87	2100
BMI-0067	444	93.6	29.68	596	164	9595	10.50	94.74	0.91	0.13	14.38	42.07	1950
BMI-0068	432	94.9	23.18	528	144	9810	10.80	96.35	0.82	0.13	15.67	43.12	1650
BMI-0069	449	92.6	24.25	528	144	9567	10.85	94.12	0.84	0.14	16.50	43.34	1875
BMI-0070	345	90.0	23.66	528	144	9704	10.80	96.11	0.95	0.16	16.64	43.26	1600
BMI-0071	455	96.0	21.77	528	144	9651	10.15	93.93	0.83	0.14	16.46	40.61	1650
BMI-0072	385	91.4	25.92	500	116	9800	10.30	96.03	0.90	0.13	15.01	41.31	1898
BMI-0073	342	93.3	30.49	500	116	9686	10.70	93.86	0.90	0.16	17.63	42.89	1950
BMI-0074	488	95.9	34.10	528	144	9698	9.95	93.74	1.25	0.22	18.04	39.89	1650

The amylase estimation was carried by using the fourth day old fifth instar larvae starved for 2 hours and kept in a clean container with dried tobacco leaves on the plastic paper over the mulberry leaves. After 20 minutes, regurgitated digestive juice was collected in syringes and stored the sample at  $-20^{\circ}\text{C}$  to avoid oxidation. Centrifuged at 5000 rpm in  $4^{\circ}\text{C}$  to get clear solution, this is used for further analysis. DNS was added to give colour (reddish orange) and to stop the reaction of enzyme over the substrate.  $10\ \mu\text{l}$  of digestive juice was taken in test tubes and 2ml of 0.2 per cent starch solution was added and incubated at  $37^{\circ}\text{C}$  for 60 minutes. Then 2ml of DNSA solution was added to each tube and kept in hot water bath 5 minutes. The optical density (OD) values were measured at 525nm (Manickam and Sadasivam, 1992) [20].

Further to determine concentration of maltose, serial dilutions were prepared from the stock solution of maltose from  $100\ \mu\text{l}$  upto 2ml. For each sample of 2ml, 2ml of DNSA was added and kept for 5 minutes in water bath. After cooling, recorded the OD values of the serially diluted maltose solutions at 525nm. A standard graph was prepared by plotting OD values against the serial dilutions. The amylase activity was calculated using the standard graph and expressed as  $\mu\text{g}/100\text{ml}$  of digestive juice in 60 minutes. The amylase activity values obtained were correlated with the growth, productive and economic traits (Table-3) using SPSS statistical packages. The amylase quantity for all the 73 MV accessions studied were represented graphically (Fig 1-3).

**Table 3:** Correlation of growth and economic traits with digestive amylase activity

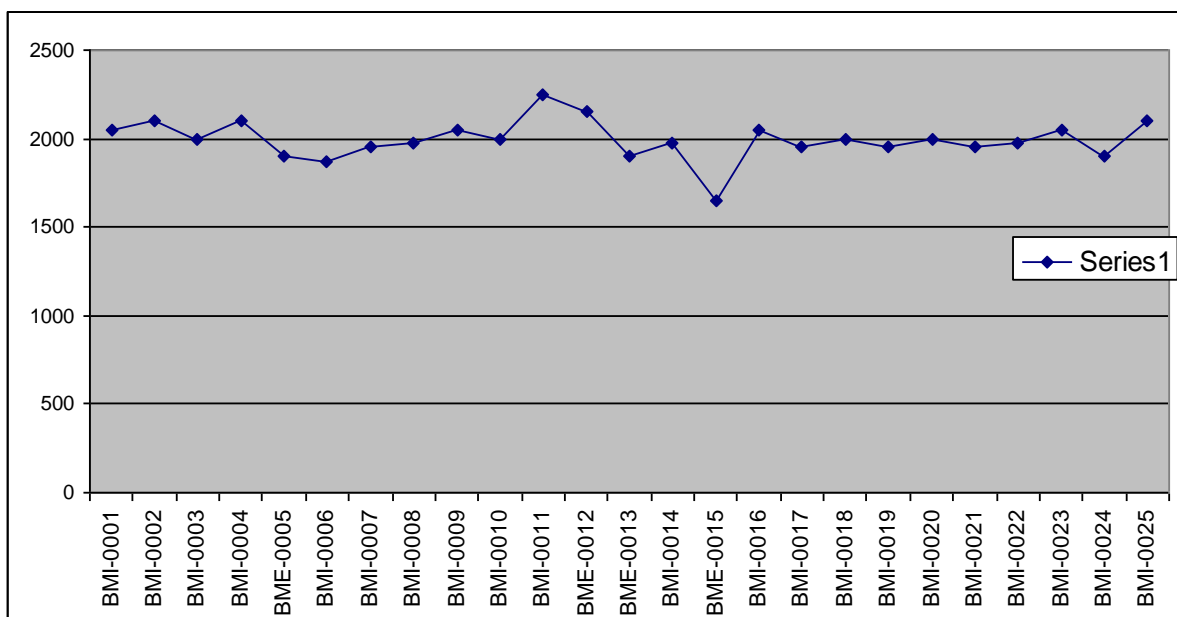
	Fecundity	Hatching	Weight of 10 larvae	Total Larval duration	Fifth age larval duration	ERR by No	ERR by Wt.	Pupation	Single cocoon weight	Single shell weight	Shell Ratio	YIELD/ 100 dfls	Amylase activity mg/ml/h
Fecundity	1												
Hatching	0.408**	1											
Weight of 10 larvae	0.129	0.012	1										
Total Larval duration	0.210	0.248*	-0.383**	1									
Fifth age larval duration	0.197	0.322**	-0.272*	0.858**	1								
ERR by No.	-0.020	-0.068	-0.150	-0.057	-0.065	1							
ERR by Wt.	0.005	-0.037	0.030	0.147	0.129	-0.114	1						
Pupation	0.011	-0.170	-0.139	-0.061	-0.026	0.870**	-0.036	1					
Single cocoon weight	0.261*	0.168	0.465**	0.058	0.122	-0.050	0.133	-0.086	1				
Single shell weight	0.217	0.198	0.442**	0.185	0.312**	-0.043	0.147	-0.113	0.808**	1			
Shell Ratio	0.077	0.144	0.219	0.232*	0.379**	-0.033	0.101	-0.107	0.246*	0.762**	1		
Yield/100 dfls	0.035	-0.068	-0.046	0.113	0.073	-0.011	0.576**	0.028	0.092	0.148	0.157	1	
Amylase activity mg/ml/h	-0.280*	-0.308**	0.135	-0.204	-0.355**	-0.099	0.070	-0.087	0.165	-0.067	-0.282*	0.004	1

\*\* Significant at 0.01 level (2 tailed) P = 0.01 n = 73

**3. Results and Discussion**

The productive and economic traits of the 73 multivoltine accessions revealed good variability. The fecundity varied from 520 (BMI-024) to 295(BMI-016) numbers/ dfl with hatching of 96% (BMI-071) to 86.9 % (BMI-053). The weight of 10 grown larvae was high 34.1 g in accession BMI-074 and it was low in BMI-042 (20.5g). Accession BMI-001 had the longest total larval duration (596 h) and BMI-034 (486h.) had the least larval duration. The fifth age larval duration was longer (164 h) in accessions BMI-067 and it was shorter in accession BMI-021 (96 h.). The effective rate of rearing by number was high in accession BMI-027 (9876) and it was low in BMI-002 (8679). The effective rate of rearing by weight was higher 12.5 kg in accession BMI-007

whereas it was low in accession BMI-065 with 8.5 kg. Accession BMI-027 showed the highest pupation (97.3%) and it was low in accession BMI-002 (85.1 %). The cocoon weight was higher in accession BMI-074 (1.25g.) and it was low in accession BMI-031 (0.76g.). The shell weight (0.22g.) and shell ratio (18.04 %) were higher in accession BMI-074 and they were low in accession BMI-037 with 0.10g and 11.04 % respectively. The cocoon yield per 100 dfls was higher in accession BMI-007 with 49.8 kg and it was low in accession BMI-009 (22.3). The amylase quantity varied from 2250 in µg /10µl /hour in accession BMI-011 and BMI-027 to 1600 µg /10µl /hour in accession BMI-074. The variability of the amylase quantity among the 73 accessions studied was graphically represented in Fig: 1-3.



**Fig 1:** Amylase activity of MV accessions (1-25 Nos.).



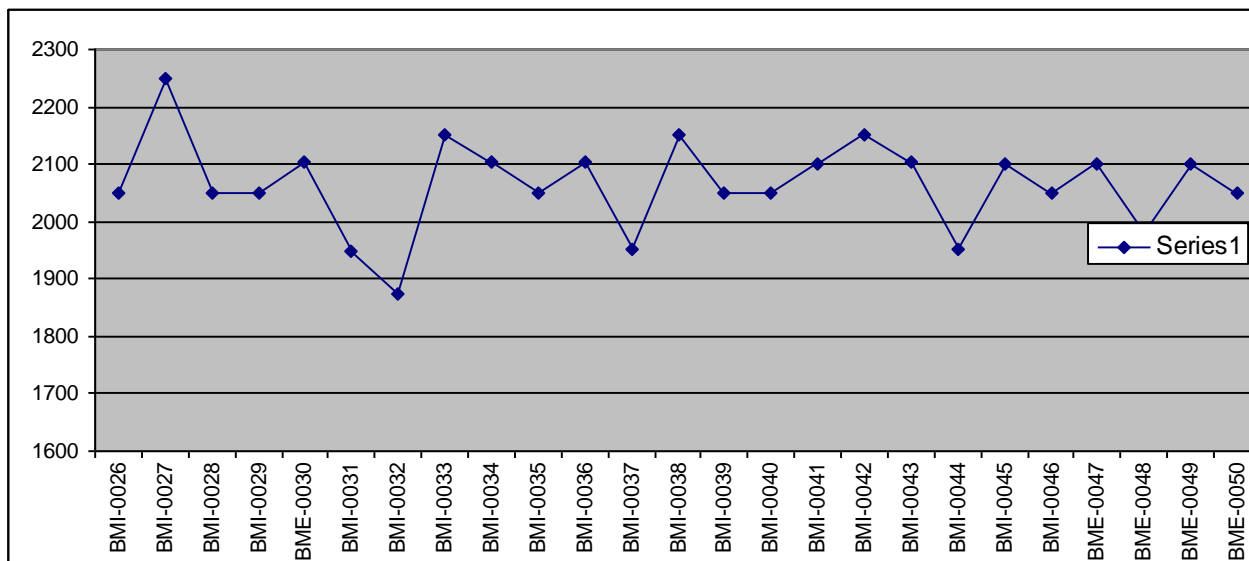


Fig 2: Amylase activity of MV accessions (26-50 Nos.)

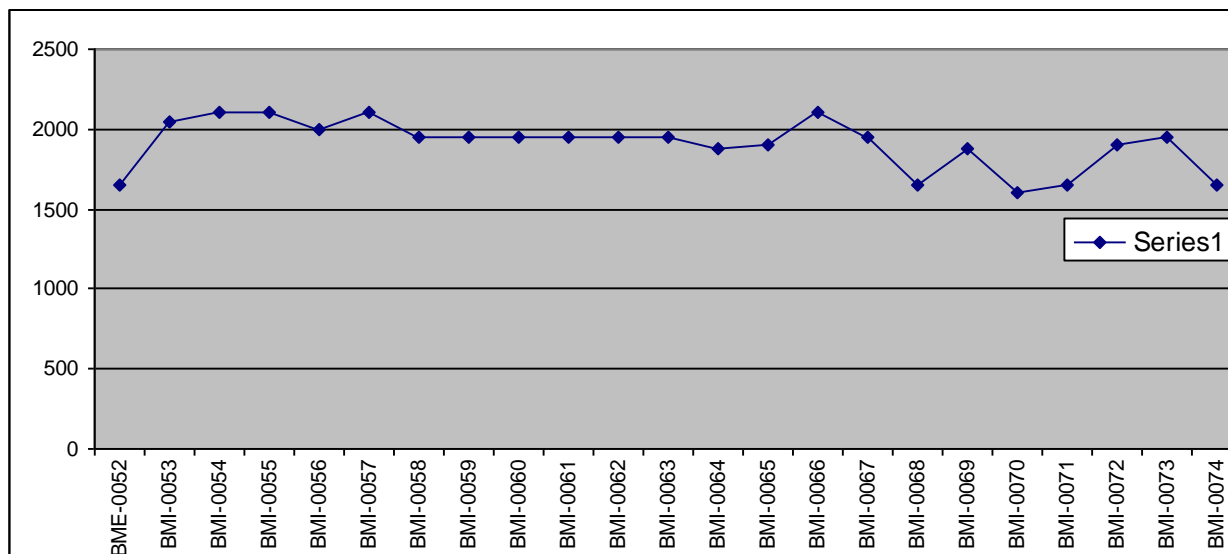


Fig 3: Amylase activity of MV accessions (52-74 Nos.).

The digestive amylase quantity was estimated among the 73 multivoltine accessions studied varied from a highest value of 2250 µg /10µl /hour to a lowest value of 1600 µg /10µl /hour by following the standard protocols. There is quite a high amount of variability among the accessions studied. The highest values of amylase activity was recorded in the accessions BMI-0011 and BMI-0027 (2250 µg /10µl /hour) followed by BME-0012, BMI-0033, BMI-0038, BMI-0042 (2150 µg /10µl /hour). Accessions BME-0030, BMI-002, BMI-0025, BMI-0034, BMI-0036 and BMI-0043 (2103 µg /10µl /hour). Nine accessions had 2100 µg /10µl /hour amylase activity viz., BME-0047, BME-0049, BMI-0004, BMI-0041, BMI-0045, BMI-0054, BMI-0055, BMI-0057 and BMI-0066. Accession BMI-0016 had 2053 µg /10µl /hour of amylase activity followed by twelve accessions BME-0050, BMI-001, BMI-0009, BMI-0023, BMI-0026, BMI-0028, BMI-0029, BMI-0035, BMI-0039, BMI-0040, BMI-0046 and BMI-0053. Five accessions BMI-0003, BMI-0010, BMI-0018, BMI-0020 and BMI-0056 showed 2000 µg /10µl /hour of amylase activity. The results observed are in concurrence

with the results of differential digestive amylase activity among the silkworm strains recorded earlier by Hirata (1971) [12].

The multivoltine accession BMI-0070 showed the least amylase activity of 1600 µg /10µl /hour followed by five accessions BMI-0074, BMI-0071, BMI-0068, BME-0052 and BME-0015. Accessions BMI-0069, BMI-0064, BMI-0032, BMI-0006 revealed amylase activity of 1875 µg /10µl /hour and it was 1898 µg /10µl /hour in the case of accession BMI-0072. Similarly 23 accessions were in the range of 1980 to 2000 µg /10µl /hour of amylase activity. Accession BME-0048 showed 1980 µg /10µl /hour followed by three accessions BMI-008, BMI-0014, BMI-0022 with amylase activity values of 1975 µg /10µl /hour and then by four more accessions BMI-0021, BMI-0037, BMI-0044 and BMI-0062. The activity of amylase was 1950 µg /10µl /hour in eleven accessions BMI-0007, BMI-0017, BMI-0019, BMI-0031, BMI-0058, BMI-0059, BMI-0060, BMI-0061, BMI-0063, BMI-0067 and BMI-0073 followed by four accessions BME-0005, BME-0013, BMI-0024 and BMI-0065.

The relation between digestive amylase activity and economic traits among the silkworm races have been reported (Hirata, 1971) <sup>[13]</sup>. Some of the ruling hardy popular multivoltine breeds like Pure Mysore (BMI-0001), Sarupat (BMI-0002), Moria (BMI-0003) and Nistari (BMI-0017) were in the high digestive amylase activity category 2050, 2103, 2000, and 1950  $\mu\text{g} / 10\mu\text{l} / \text{hour}$  respectively. The high yielding accessions BMI-0070 and four accessions BMI-074 BMI-0081, BME-0052 and BME-0015 showed very least quantity of amylase activity 1600, 1650  $\mu\text{g} / 10\mu\text{l} / \text{hour}$  respectively where as some of the low yielding accessions like BMI-027 and BMI-011 had highest quantity of amylase of 2250  $\mu\text{g} / 10\mu\text{l} / \text{hour}$  which is in concurrence with the earlier studies conducted by Chatterjee *et al.*, (1992) <sup>[9]</sup> and Patnaik *et al.*, (1995) <sup>[22]</sup>. These accessions could be used as resource materials for inclusion in breeding programmes oriented towards amylase specific breeds and also in hard and robust breeds for the tropical regions to improve the silk productivity.

Correlation between amylase activity in 73 multivoltine silkworm genetic resources with that of the 12 productive traits (table-3) revealed positive and non-significant correlations between weight of larvae (g), ERR by weight (kg.) and single cocoon weight (g) and yield/100dfls (kg.) where as non-significant negative correlations were observed in the traits like total larval duration (h), ERR by number (no.) pupation rate (%) and single shell weight (g). There were negative and significant correlations at 5 % level in the case of fecundity (no.) and shell ratio (%) and significant negative correlations at 1 % level was observed in the traits like hatching (%), fifth age larval duration (h). The studies in line with that of the earlier studies by Chatterjee *et al.* (1988; 1989; 1990; 1993) <sup>[6-10]</sup>.

The results obtained indicated there is much variability in the quantum of amylase recorded among the 73 multivoltine accessions studied as per the standard procedures which is in line with the earlier studies on the importance of amylase *viz.*, Chatterjee *et al.*, (1992) <sup>[9]</sup>; Abraham *et al.*, (1992) <sup>[1]</sup>; Patnaik and Datta, (1995) <sup>[22]</sup>.

Datta and Ashwath (2000) <sup>[11]</sup> reported that the prospects of using digestive amylase as a marker for improving the viability in silkworm as it contributes for better digestibility and has close association with survival. Therefore the multivoltine accessions showing higher digestive amylase activity could be used as breeding resource materials for developing hardy breeds which could have higher abiotic stress tolerance and can digest even the mediocre/inferior quality of leaves which are bound to available in the not so conducive seasons also by withstanding stress conditions and higher survival.

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